Purification His-tagged proteins (written by Zhenhao Fang, Ikura Lab)

- Ni-NTA resin is stored at 4 degrees as 50% slurry in 20% ethanol
- Centrifuge for 2 min. at 700 xg
- Remove supernatant
- Add 2x resin-bed volume of His Wash buffer 2 centrifuge for 2 min at 700 xg, remove supernatant and repeat
- Resuspend resin with the protein lysis supernatant
- Stir 100 ml lysis solution with 7 ml Ni-resin (not slurry) in a 250 ml glass beaker in the cold room for 30 min, under gently magnetic stirring (= batch binding)
- Wet the filter in the yellow glass column with tap water
- Pour supernatant and resin to the column, collect the flow through
- Take 10 ul flow through for SDS-PAGE
- Wash with 20 column volumes (150 ml) of wash buffer 1 and 10 column volumes (75 ml) of wash buffer 2.
- Use Bradford to check if wash buffers don't contain protein anymore
- Take out 10 ul wash buffer and 10 ul resin for SDS-PAGE
- Add elution buffer (contains 250 mM imidazole)
- Save elution steps in eppendorf tubes ($25x \sim 1.5$ ml). Protein elutes in 40 ml
- Take 10 ul of elution for SDS-PAGE
- Pool steps that contain protein (~2 18)
- Resin can be used at least 5x before regeneration
- Wash resin with 10x C.V. MES 20 mM pH 5, 100 mM NaCl and 10x C.V MQ water.
- Store as 50% slurry in 20% ethanol in cold room

For uncleaved Ras

- Buffer exchange to reduce imidazole amount
- Use Millipore 10K centrifugal filter 4000 xg for 15 min. to concentrate (to ~ 5ml). Add wash buffer 2 and mix! Repeat 1x. Concentrate to < 10 ml (= max for gel filtration chromatography)
- Sample to Akta (S75 column)

To cleave His-tag Ras

- Concentrate to ~10-15 ml
- Add 50 uM GDP & 10 ul thrombin (scales with cell culture volume)
- Dialyse overnight in 4L general dialysis buffer
- Take out 20 ul for SDS-PAGE sample
- Transfer to 50 ml falcon
- Add 15 ul thrombin, leave at RT for 3h (gently invert every 30-60 min)
- Put protein on ice
- Run SDS-PAGE to confirm cleaving of the His-tag, otherwise leave at RT for another hour
- Concentrate to < 10 ml
- Sample to Akta (S75 column)

• His Wash buffer 1Liter	
Tris pH 8, 50 mM	50 ml from 1 M stock
NaCl, 500 mM	166.7 ml from 3M stock
Glycerol, 10%	200 ml of 50% stock
Imidazole, 10 mM	10 ml from 1 M stock pH 8
MgCl2, 5 mM	5 mL from 1 M stock
Add b-ME 10 mM	x mL from 14.3 M stock just before use

• His wash buffer 0.5 Liter Tris pH 8, 50 mM NaCl, 150 mM Glycerol, 10% Imidazole, 10 mM MgCl2, 5 mM Add b-ME, 10 mM

25 ml from 1 M stock 25 ml from 3 M stock 100 ml from 50% stock 5 ml from 1 M stock 2.5 ml from 1 M stock x mL from 14.3 stock just before use

His Elution buffer 0.5 Liter
Tris pH 8, 50 mM
NaCl, 150 mM
Glycerol, 10%
Imidazole, 250 mM
MgCl2, 5 mM
Add b-ME 10 mM
25 ml from 1 M stock
x mL from 14.3 M stock just before use

Kras gel filtration buffer 1 liter HEPES, pH 7.4, 20 mM 20 NaCl, 100 mM 33 MgCl2, 5 mM 5 TCEP 2 mM 0. TCEP reduces the pH Add up to 750 mL MQ water Ajust pH to 7.4 Fill to 1L

20 ml from 1 M stock 33.33 mL from 3 M stock 5 ml from 1 M stock 0.5733 gram (Mw = 286.65)

• General dialysis buffer 1 lite	r
Tris pH 8, 50 mM	50 ml from 1M stock
NaCl, 150 mM	50 ml from 3 M stock
Glycerol, 2%	40 ml from 50% stock
MgCl2, 5 mM	5 ml from 1 M stock
DTT, 1 mM	1 ml from 1M stock (add just before starting dialysis)